

# The H90-10 single-chain antibody recognizes Hsp90 $\beta$ by immunoprecipitation and Western blotting

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## Abstract

The recombinant antibody H90-10 detects the endogenous human heat-shock protein 90 beta (Hsp90 $\beta$ ) by immunoprecipitation (IP) and Western blotting.

## Introduction

The mouse monoclonal antibody H90-10 specifically recognizes the Hsp90 $\beta$  (UniProt #P08238) isoform of the Hsp90 family (Holt *et al.*, 1999; Barent *et al.*, 1998). Here, we describe the ability of the single-chain variable antibody (scFv) H90-10 to immunoprecipitate the endogenous human Hsp90 $\beta$  and to recognize it by immunoblotting.

## Materials & Methods

**Antibodies:** The ABCD\_AO870 antibody (ABCD nomenclature, <https://web.expsy.org/abcd>) was produced by the Geneva Antibody Facility (<https://www.unige.ch/medecine/antibodies>) as a mini-antibody with the antigen-binding scFv fused to the Fc region of mouse IgG2a. The synthesized scFv sequence (GeneArt, Invitrogen) corresponds to the sequence of the variable regions of the anti-Hsp90 $\beta$  monoclonal H90-10 (Holt *et al.*, 1999; Barent *et al.*, 1998) joined by a peptide linker (GGGGS)<sub>3</sub>. The H90-10 variable sequences were determined with permission by Brian C. Freeman (University of Illinois, Urbana) from the H90-10 hybridoma originally from David O. Toft (Mayo Clinic, Rochester). HEK293 suspension cells (growing in FreeStyle™ 293 Expression Medium, Gibco #12338) were transiently transfected with the vector coding for the scFv-Fc. Supernatant (~90 mg/L) was collected after 5 days.

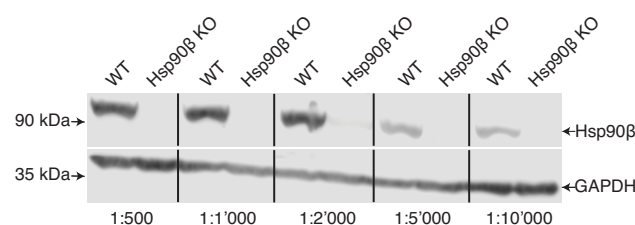
**Antigen:** Both wild-type HEK293T cells, which endogenously express Hsp90 $\beta$ , and their Hsp90 $\beta$  knock-out counterpart (Bhattacharya *et al.*, 2020) were grown in Dulbecco's Modified Eagle's Medium supplemented with GlutaMAX, 10% fetal bovine serum, and penicillin/streptomycin (100 U/ml).

**Protocol:** Cells were pelleted and lysed in lysis buffer (10 mM Tris-HCl pH 7.5, 1 mM EDTA, 10 mM NaCl, 10 mM Na-molybdate, 10% glycerol, 1 mM DTT, 0.1% Triton X-100) with 1x protease inhibitor complex for 50 min at 4 °C using a Bioruptur™ Twin sonicator. Extracts were centrifuged at 16'100 g for 10 min at 4 °C and the pellets were discarded. 500  $\mu$ g of proteins were used for the IP. Samples were incubated overnight with murine IgG (Sigma-Aldrich #I5381) as negative control, H90-10, or scFv H90-10, diluted at 1:250, 1:50, and 1:50, respectively. The next day, 50  $\mu$ l of Dynabeads™ Protein

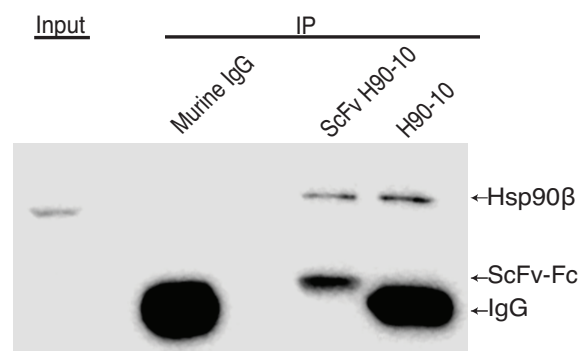
G (Invitrogen #10009D) were added for three hours and then washed 6 times for 10 min with lysis buffer. 50  $\mu$ g extract (Fig. 1), and 20  $\mu$ g input extract and IP samples (Fig. 2) were loaded on separate 10% SDS-polyacrylamide gel and then transferred to a nitrocellulose membrane (100 V, 105 min). The membranes were blocked for 60 min with 5% w/v non-fat dry milk in Tris-buffered saline containing 0.2% Tween 20 (TBST), then incubated overnight at 4 °C with the different antibody dilutions in TBST. As a loading control, corresponding sections of the same membranes were probed with an anti-GAPDH antibody (Hyttest, #5G4, dilution 1:5'000). After washing the membranes three times for 15 min with TBST, they were incubated for 90 min at room temperature with horseradish peroxidase-coupled goat anti-mouse antibody (Invitrogen #31430, dilution 1:10'000 in TBST) and washed again three times for 15 min. The immunoblot of the IP experiment was probed similarly as indicated. Chemiluminescent signals were recorded with a LI-COR Odyssey Fc Imaging System.

## Results

The scFv version of H90-10 specifically recognizes Hsp90 $\beta$  (Fig. 1), and it immunoprecipitates Hsp90 $\beta$  as well as or better than the original monoclonal H90-10 (Fig. 2).



**Fig. 1.** Immunoblot showing specific recognition of Hsp90 $\beta$  by the scFv H90-10 antibody at different dilutions. Extracts of Hsp90 $\beta$  KO cells were used as the negative control and GAPDH as the loading control.



**Fig. 2.** Immunoblot of an IP experiment comparing the original monoclonal H90-10 and the scFv version, with normal murine IgG as negative control, probed with scFv H90-10 (dilution 1:1'000).

## References

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## Conflict of interest

The authors declare no conflict of interest.