

The AE765, AK423, AK692, AO233 and AO234 antibodies recognize human actin by immunofluorescence in HEK cells

Anna Marchetti

Geneva Antibody Facility, Faculty of Medicine, University of Geneva, 1 rue Michel Servet, CH-1211, Geneva, Switzerland

Abstract

The AE765, AK692, AO233 and AO234 antibodies recognize human actin in PAF-fixed HEK cells by immunofluorescence. Phalloidin, which recognizes polymerized actin, is more adequate to label actin filamentous structures. The AK423 antibody recognizes human actin in methanol-fixed HEK cells, but in these conditions the actin network is poorly preserved.

Introduction

Actin is one of the most abundant proteins in eukaryotic cells, and a major structural component of the cytoskeleton, forming networks of microfilaments in the cytoplasm of cells. The human genome contains six genes for actin (three for α -actin, one for β -actin, and two for γ -actin) (Pollard, 2016). Five recombinant antibodies (AE765, AK423, AK692, AO233 and AO234) were tested for their ability to label actin by immunofluorescence.

Materials & Methods

Antibodies: ABCD_AE765, ABCD_AK423, ABCD_AK692, ABCD_AO233 and ABCD_AO234 antibodies (<https://web.expasy.org/abcd/>, ABCD nomenclature) were produced by the Geneva Antibody Facility (<https://www.unige.ch/medecine/antibodies/>) as mini-antibodies with the antigen-binding scFv fused to a rabbit IgG Fc. The synthesized scFv or VHH sequences (GeneArt, Invitrogen) correspond to the sequences of the variable regions joined by a peptide linker (GGGS)₃ (Table 1). HEK293 suspension cells (growing in HEK TF medium, Xcell 861-0001, supplemented with 0.1% Pluronic F68, Sigma P1300) were transiently transfected with the vector coding for the scFv-Fc or VHH-Fc. Supernatants (see Table 1 for individual yields) were collected after 4 days.

The antibody AJ519, which recognizes the TAC antigen (human IL2RA, Uniprot P01589), was used as a negative control (Arsimoles *et al.*, 2020). As positive control, a commercial anti-beta actin antibody (clone 2D4H5, Proteintech 66009-1-Ig), raised against human ACTB (Uniprot P60709), was used.

Table 1: Clone number, epitope, reference and production yields for the antibodies used in this study.

ABCD	Clone	Epitope	Reference	Yield (mg/L)
AE765	SA1A	Human actin and alpha-actinin	Victor <i>et al.</i> , 1992	70
AK423	mAb 236	<i>Dictyostelium</i> actin (Uniprot P07830)	Lima, 2019	10
AK692	Nb141	Human ACTB, (Uniprot P60709)	Jovčevska <i>et al.</i> , 2014	<5
AO233	3-1		Persson <i>et al.</i> , 2013	90
AO234	3-2			30

Antigen: HEK cells were cultured on glass coverslips (Menzel-Gläser, 22x22 mm) and grown in DMEM GlutaMAX™ (Gibco 31966) supplemented with 8% Fetal Bovine Serum (Gibco 10270).

Protocol: The whole procedure was carried out at room temperature. Cells were rinsed once with PBS, and fixed either with (i) PBS + 4% paraformaldehyde (PAF) (w/v) (Applichem A3013) for 30 min, blocked with PBS + 40 mM ammonium chloride (NH₄Cl) (Applichem A3661) for 5 min, and then permeabilized in PBS + 0.2% saponin for 5 min; or (ii) methanol at -20 °C for 2 min. Fixed cells were washed once (5 min) in PBS and once with PBS + 0.2% (w/v) BSA (PBS-BSA), and incubated for 30 min with the primary antibodies (for the ABCD antibodies, final concentration 5 mg/L in PBS-Tween; for the Proteintech antibody, 0.05 mg/L). After 3 washes (10 min) with PBS-BSA, cells were incubated for 30 min with secondary goat anti-rabbit IgG conjugated to AlexaFluor-647 (1:400, Molecular Probes A21245, for the ABCD antibodies) or anti-mouse IgG conjugated to AlexaFluor-647 (1:400, Molecular Probes A21235, for the Proteintech antibody). After 3 washes (10 min) with PBS-BSA, cells were incubated for 30 min with Phalloidin-TRITC (1 μ g/ml in PBS-BSA, Sigma P1951). After 3 washes (10 min) with PBS-BSA, cells were mounted on slides (Menzel-Gläser, 76x26 mm) with M \ddot{u} wiöl (Hoechst) + 2.5% (w/v) DABCO (Fluka 33480). Pictures were taken using a Zeiss LSM700 confocal microscope, with a 63x Neofluar oil immersion objective.

Results

Fluorescent phalloidin was used as a marker to label polymerized actin filaments in PAF-fixed HEK cells (Fig. 1, in white). Four anti-actin antibodies, AE765, AK692, AO233 and AO234, label some of the same structures stained with phalloidin (Fig. 1, in cyan; zoomed-in regions, identified by arrows, can be seen in Fig. 2). They also label the cellular cytoplasm (Fig. 2, pinheads). Although this cytosolic staining may represent a background staining, it seems more likely that it is due to detection of non-polymerized actin.

No staining was observed when the primary anti-TAC antibody was used as a negative control (Fig. 1, 'Neg ctr'). In PAF-fixed HEK cells, no staining was observed with the commercial anti-actin 2D4H5 antibody, nor with AK423 (Figs. 1 and 2).

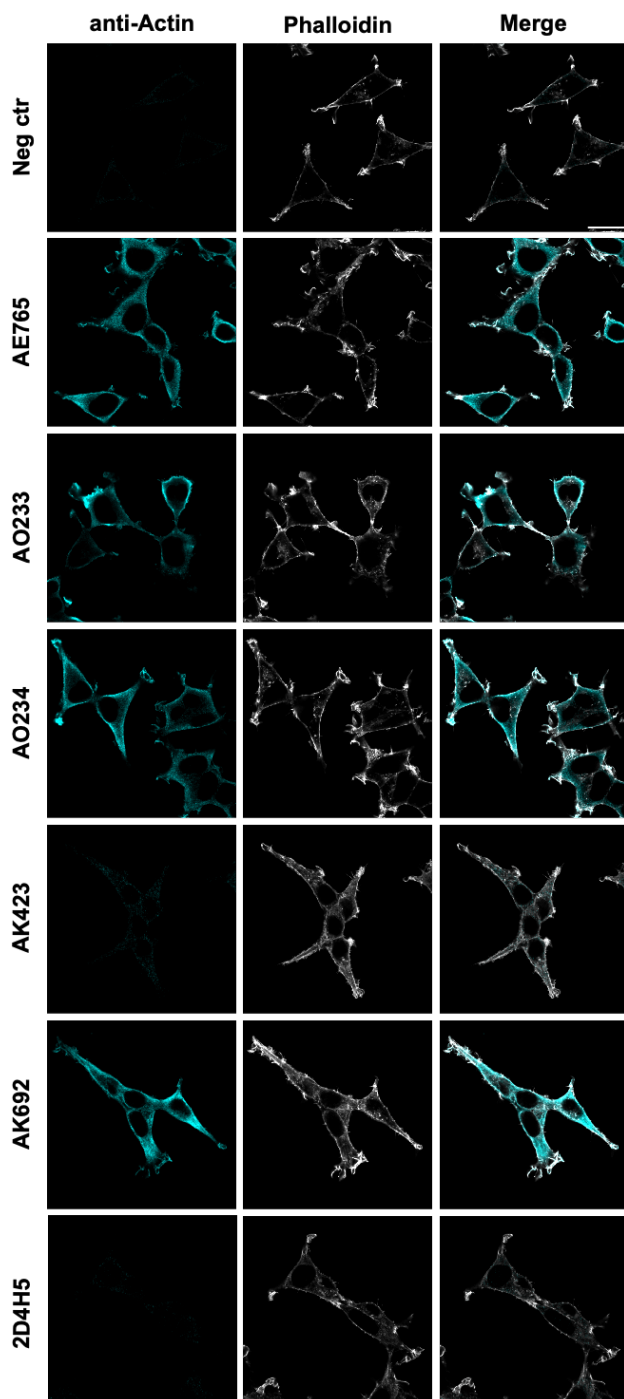


Fig. 1. Actin labelling in PAF-fixed HEK cells. Labeling with AE765, AK692, AO233 and AO234 antibodies ('anti-Actin', in cyan) partially co-localizes with phalloidin staining ('Phalloidin', in white). Labeling of cytoplasmic structures not labelled by phalloidin was also seen. No labelling was observed when an unrelated primary antibody (anti-TAC AJ519) was used ('Neg ctr' panel). No labelling was seen with AK423 or the commercial 2D4H5 antibodies. Scale bar: 20 μ m.

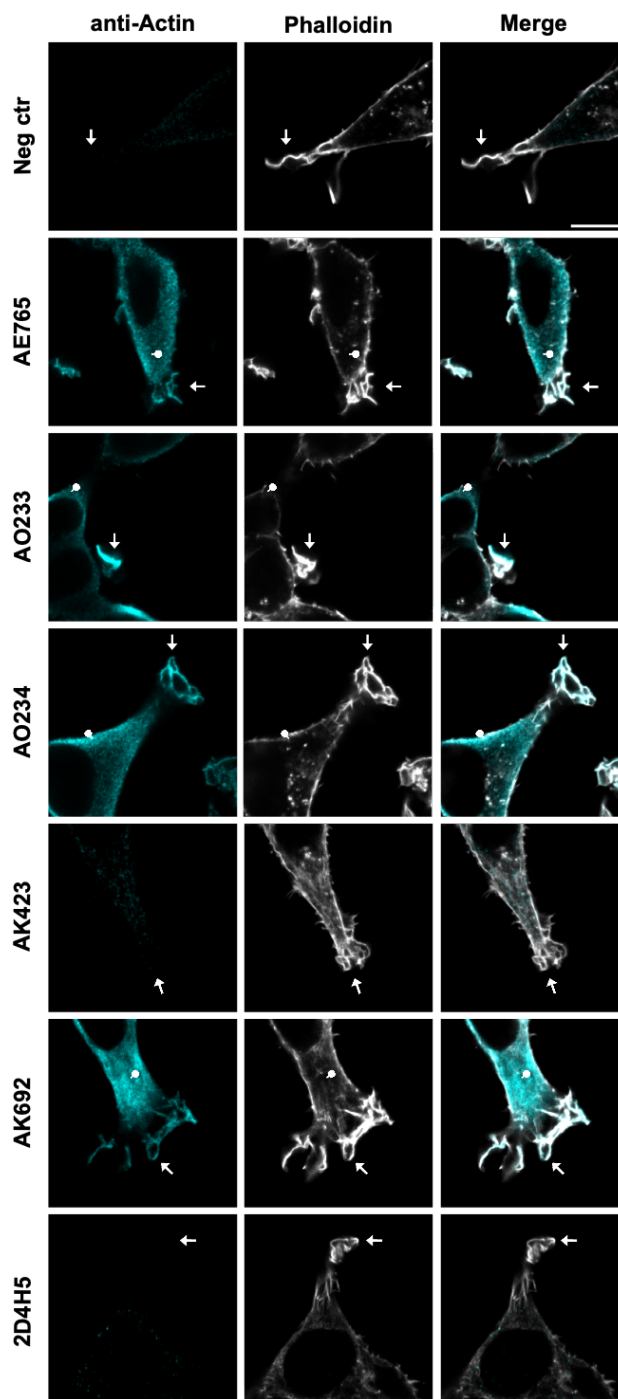


Fig. 2. Actin labelling in PAF-fixed HEK cells. Insets from Fig. 1, showing partial co-localization of AE765, AK692, AO233 and AO234 with phalloidin (arrows) and the labelling of cytoplasmic structures not labelled by phalloidin (pinheads). Scale bar: 10 μ m.

Methanol fixation abrogates the signal of phalloidin (as it destroys the native conformation of the F-actin; Mierke, 2018). In methanol-fixed HEK cells, AE765, AK692, AO233 and AO234 did not label any recognizable structures (Fig. 3).

On the contrary, both the commercial anti-actin 2D4H5 and AK423 antibodies label structures that likely represent poorly conserved filamentous actin structures (Fig. 3, arrows).

References

Arsimoles D, D'Esposito AG, Gaspoz V, *et al.* The AJ519 antibody labels the human TAC/IL2RA protein by immunofluorescence. *Antibody Reports*, 2020, 3:e118. doi:10.22450/journals/abrep.2020.e118

Jovčevska I, Zupanec N, Kočevnar N, *et al.* TRIM28 and β -actin identified via nanobody-based reverse proteomics approach as possible human glioblastoma biomarkers. *PLoS One*. 2014; 9(11):e113688. PMID: 25419715.

Lima WC. The AK423 antibody recognizes *Dictyostelium* actin network by immunofluorescence. *Antibody Reports*, 2019, 2:e54. doi:10.22450/journals/abrep.2019.e54

Mierke CT. Role of the actin cytoskeleton during matrix invasion. In *Physics of Cancer*, v. 1, 2018, 7:1-87. doi: 10.1088/978-0-7503-1753-5ch7

Persson H, Ye W, Wernimont A, *et al.* CDR-H3 diversity is not required for antigen recognition by synthetic antibodies. *J Mol Biol*. 2013; 425(4):803-11. PMID: 23219464

Pollard TD. Actin and actin-binding proteins. *Cold Spring Harb Perspect Biol*. 2016; 8(8):a018226. PMID: 26988969.

Victor KD, Pascual V, Williams CL, Lennon VA, Capra JD. Human monoclonal striational autoantibodies isolated from thymic B lymphocytes of patients with myasthenia gravis use VH and VL gene segments associated with the autoimmune repertoire. *Eur J Immunol*. 1992; 22(9):2231-6. PMID: 1516616.

Conflict of interest

The authors declare no conflict of interest.

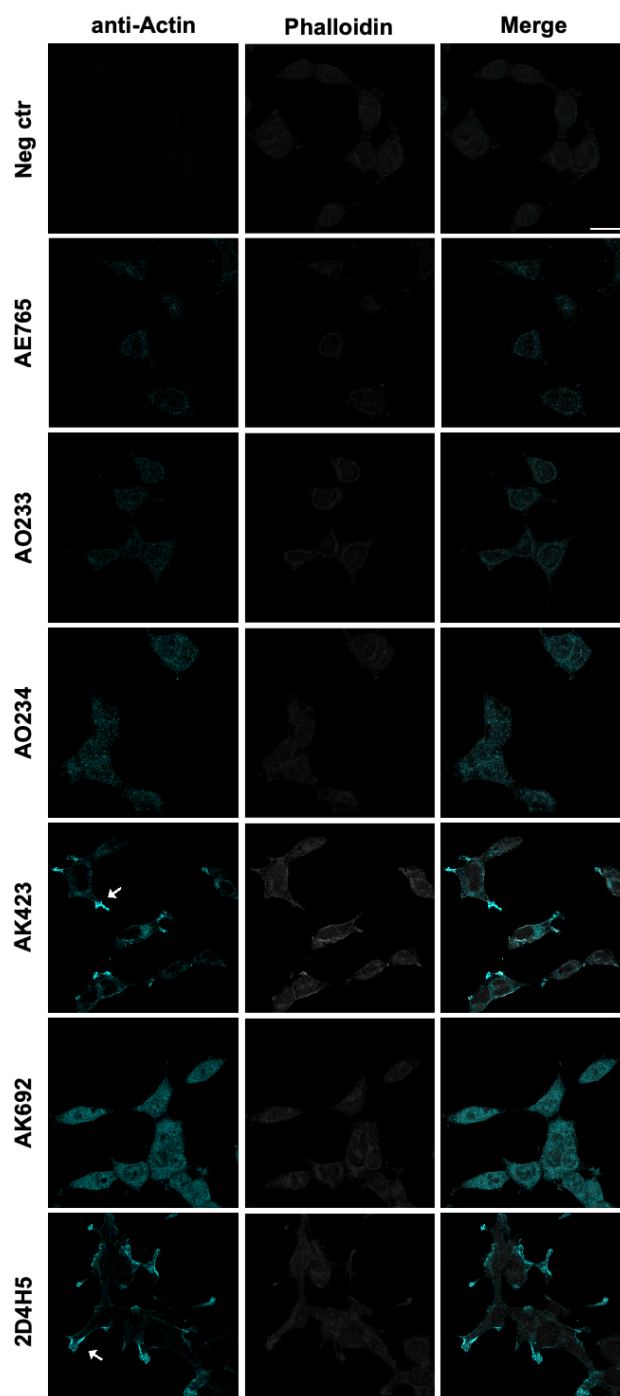


Fig. 3. Actin labelling in methanol-fixed HEK cells. No phalloidin staining was seen in methanol-fixed cells ('Phalloidin', in white). Labeling with AE765, AK692, AO233 and AO234 antibodies ('anti-Actin', in cyan) was also abrogated. No labelling was seen when an unrelated primary antibody (anti-TAC AJ519) was used ('Neg ctr' panel). AK423 and the commercial 2D4H5 antibodies label actin structures (arrows). Scale bar: 20 μ m.