RB388, RB389, RB390, RB391 and RB392 antibodies do not recognize the Dictyostelium AlyL protein by Western blot

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Abstract

The recombinant antibodies RB388, RB389, RB390, RB391 and RB392 do not detect by Western blot the full-length AlyL protein from *Dictyostelium discoideum*.

Introduction

AlyL (Amoeba LYsozyme Like, DDB_G0286229, UniProt #Q54M35) is a member of the amoeba lysozyme family in the amoeba *D. discoideum*. Here we describe the inability of five recombinant antibodies (RB388, RB389, RB390, RB391 and RB392) to detect the full-length AlyL protein by Western blot.

Materials & Methods

Antibodies: ABCD_RB388, ABCD_RB389, ABCD_RB390, ABCD_RB391 and ABCD_RB392 antibodies (ABCD nomenclature, https://web.expasy.org/abcd/) were produced by the Geneva Antibody Facility (www.unige.ch/medecine/antibodies; Blanc *et al.*, 2014) as mini-antibodies with the antigen-binding scFv fused to a mouse Fc (MRB388, MRB389, MRB390, MRB391 and MRB392). HEK293 suspension cells (growing in FreeStyleTM 293 Expression Medium, Gibco #12338) were transiently transfected with the vector coding for the scFv-Fc of each antibody. Supernatants (~50 mg/L) were collected after 5 days. As a positive control, the anti-6xHis antibody AD946 (Lamrabet and Jauslin, 2018) was used.

Antigen: The antibodies were raised against a N-biotinylated synthetic peptide corresponding to 23 residues close to AlyL C-terminus (RVISTNVGPNV DIEDKIGKPIMD). *D. discoideum* DH1 (WT) cells expressing a 6xHis-tagged AlyL protein (AlyL-His, 6xHis-tag fused to the C-terminus) were used to detect the full-length AlyL protein. *AlyL* knockout (KO) cells were used as a negative control.

Protocol: 5x10⁶ *D. discoideum* cells were pelleted and resuspended in 200 μL of reducing sample buffer (20.6% (w/v) sucrose, 100 mM Tris pH 6.8, 10 mM EDTA, 0.1% (w/v) bromophenol blue, 4% (w/v) SDS, 6% (v/v) β-mercaptoethanol). 20 μL of each sample was migrated (200 V, 30 min) in a 10% acrylamide gel (Mini-PROTEAN® TGXTM Precast Gel, Biorad #456-1033), and transferred to a nitrocellulose membrane using a dry transfer system for 10 minutes (iBlot gel transfer device, Invitrogen #IB1001EU). The membranes were blocked during 1 hour in PBS containing 0.1% (v/v) Tween20 and

7% (w/v) milk, and washed three times for 15 minutes in PBS + 0.1% (v/v) Tween20. The membranes were then incubated with each of the six tested antibodies (dilution 1:10 and 1:2 in PBS-Tween for MRBs and AD946, respectively), overnight at 4 °C, then washed three times for 15 minutes. The membranes were then incubated with horseradish peroxidase-coupled goat anti-mouse IgG (Biorad #170-6516, dilution 1:3000) and washed twice for 15 minutes and once for 5 minutes in PBS-Tween. The signal was revealed by enhanced chemiluminescence (ECL) (Amersham Biosciences) using a PXi-4 gel imaging systems (Syngene).

Results

Antibodies MRB388, MRB389, MRB390, MRB391 and MRB392 did not specifically recognize the AlyL protein in *D. discoideum* cells overexpressing a 6xHis-tagged AlyL protein (Fig. 1). The protein could be detected with an anti-6xHis antibody (AD946): a band is detected in the overexpressing cells, but is absent on *alyL*-KO cells.

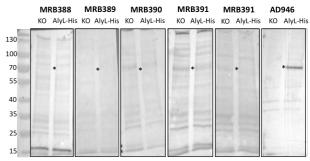


Fig. 1. No specific binding of MRB antibodies to cells overexpressing AlyL-His. AlyL-His was successfully detected by the anti-6xHis AD946 antibody (position indicated by an asterisk), but not by any of the MRB antibodies.

References

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Conflict of interest

The authors declare no conflict of interest.