

# The AJ517 antibody labels the mouse CD8 $\beta$ protein by western blot

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## Abstract

The AJ517 antibody detects the mouse CD8 $\beta$  protein by western blot.

## Introduction

CD8 is a membrane-bound glycoprotein complex expressed primarily in cytotoxic T lymphocytes. It is composed of two transmembrane subunits,  $\alpha$  and  $\beta$ , that associate to form a disulfide-linked heterodimer (Parnes, 1989) present at the cell surface. Here, we describe the ability of the AJ517 antibody, a single chain fragment (scFv) derived from the 35.17.2 hybridoma, to successfully detect the CD8 $\beta$  protein (Uniprot #P10300) by western blot in HEK293 cells expressing CD8 $\alpha$  and CD8 $\beta$ .

## Materials & Methods

**Antibodies:** ABCD\_AJ517 antibody (ABCD nomenclature, web.expasy.org/abcd/; Lima *et al.*, 2019) was produced by the Geneva Antibody Facility (www.unige.ch/antibodies/) as mini-antibody with the antigen-binding scFv fused to a rabbit IgG Fc. The synthesized scFv sequence (GeneArt, Invitrogen) correspond to the sequence of the variable regions of the 35.17.2 hybridoma (Golstein *et al.*, 1982) joined by a peptide linker (GGGG)<sub>3</sub>. The sequencing of the 35.17.2 hybridoma was performed by the Geneva Antibody Facility. HEK293 suspension cells (growing in serum-free FreeStyle™ 293 Expression Medium, Gibco #12338) were transiently transfected with the vector coding for the scFv-Fc. AJ517 supernatant (30 mg/L) was collected after 4 days.

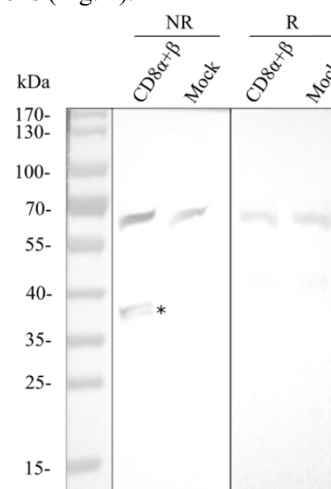
**Antigen:** The 35.17.2 hybridoma was originally raised against human thymocytes in BALB/c mice (Golstein *et al.*, 1982). HEK293 suspension cells (growing in FreeStyle™ 293 Expression Medium, Gibco #12338) were transiently transfected 3 days before the experiment with the vectors coding for the full-length mouse CD8 $\alpha$  (Uniprot #P01731) and CD8 $\beta$  protein. Co-transfection with the CD8 $\alpha$  encoding plasmid was performed to guarantee proper protein dimerization and trafficking.

**Protocol:** 5x10<sup>6</sup> transfected HEK cells were pelleted and lysed in PBS containing 0.5% (v/v) Triton X-100. Nucleus were pelleted by centrifugation (10 min at 12.000 g) and supernatant was recovered and mixed with reducing or non-reducing sample buffer (20.6% (w/v) sucrose, 100 mM Tris pH 6.8, 10 mM EDTA, 0.1% (w/v) bromophenol blue, 4% (w/v) SDS, +/- 6% (v/v)  $\beta$ -mercaptoethanol). 20  $\mu$ L of each sample was migrated (200 V, 30 min) in a 4-20% acrylamide gel (SurePAGE Bis-Tris, Genscript #M00655), and transferred to a nitrocellulose membrane using a dry transfer system for 10 minutes (iBlot gel

transfer device, Invitrogen #IB1001EU). The membranes were blocked overnight in PBS containing 0.1% (v/v) Tween20 and 7% (w/v) milk and washed three times for 5 minutes in PBS + 0.1% (v/v) Tween 20. The membranes were then incubated with the primary antibody AJ517 (dilution 1:10 in PBS-Tween) for 1 hour at room temperature and washed three times for 5 minutes. The membranes were then incubated with horseradish peroxidase-coupled goat anti-rabbit IgG (Sigma-Aldrich #A8275, dilution 1:3000) and washed twice for 5 minutes and once for 15 minutes in PBS-Tween. The signal was revealed by enhanced chemiluminescence (ECL) (Millipore) using a PXi-4 gel imaging systems (Syngene).

## Results

The antibody AJ517 recognizes the CD8 $\beta$  protein in non-reducing conditions. No signal was detected in mock transfected cells (Fig. 1).



**Fig. 1.** Specific binding of the AJ517 antibody to CD8 $\beta$  protein (position indicated by an asterisk) in cells expressing CD8 $\alpha$  and CD8 $\beta$  in non-reducing conditions (NR), but not in reducing conditions (R). No band was observed in mock transfected cells.

## References

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## Conflict of interest

The authors declare no conflict of interest.